

# Facile measurement of $^1\text{H}$ – $^{15}\text{N}$ residual dipolar couplings in larger perdeuterated proteins

Nicholas C. Fitzkee · Ad Bax

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**Abstract** We present a simple method, ARTSY, for extracting  $^1J_{\text{NH}}$  couplings and  $^1\text{H}$ – $^{15}\text{N}$  RDCs from an interleaved set of two-dimensional  $^1\text{H}$ – $^{15}\text{N}$  TROSY-HSQC spectra, based on the principle of quantitative J correlation. The primary advantage of the ARTSY method over other methods is the ability to measure couplings without scaling peak positions or altering the narrow line widths characteristic of TROSY spectra. Accuracy of the method is demonstrated for the model system GB3. Application to the catalytic core domain of HIV integrase, a 36 kDa homodimer with unfavorable spectral characteristics, demonstrates its practical utility. Precision of the RDC measurement is limited by the signal-to-noise ratio, S/N, achievable in the 2D TROSY-HSQC spectrum, and is approximately given by  $30/(S/N)$  Hz.

**Keywords** ARTSY · Catalytic core domain · HIV integrase · Quantitative J correlation · TROSY · RDC

Residual dipolar couplings (RDCs) in proteins, as measured by solution state NMR, constitute an important source of both structural and dynamic information (Prestegard et al. 2000; Bax and Grishaev 2005; Tolman and Ruan 2006). Although a wide range of different types of RDCs can be measured, including  $^1\text{H}$ – $^1\text{H}$ ,  $^1\text{H}$ – $^{13}\text{C}$ ,

$^1\text{H}$ – $^{15}\text{N}$ ,  $^{15}\text{N}$ – $^{13}\text{C}$ , and  $^{13}\text{C}$ – $^{13}\text{C}$  (Tjandra and Bax 1997; Yang et al. 1999; Permi et al. 2000; Boisbouvier et al. 2003; Vijayan and Zweckstetter 2005), in larger, slowly tumbling proteins measurements are often restricted to the backbone amide  $^1\text{H}$ – $^{15}\text{N}$  RDC,  $^1D_{\text{NH}}$ . With assignment of the backbone amide signals being a prerequisite for any detailed analysis of protein structure and dynamics, the additional effort needed to collect such couplings for modest size proteins is limited, and they can readily be obtained from the  $^{15}\text{N}$ – $\{^1\text{H}\}$  splittings in 2D or 3D NMR spectra, recorded in the absence of  $^1\text{H}$  decoupling during  $^{15}\text{N}$  evolution (Tolman et al. 1995). Increased spectral overlap associated with the doubling of the number of resonances in such spectra can be mitigated by separating the two doublet components by the IPAP method (Ottiger et al. 1998; Yao et al. 2009), or by separately recording spectra for each multiplet component using the principle of spin-state selective polarization transfer (Lerche et al. 1999).

For larger proteins, a problem associated with the frequency domain measurements is caused by the very different intrinsic line widths of the two  $^{15}\text{N}$ – $\{^1\text{H}\}$  doublet components. The downfield component benefits from favorable relaxation interference between the  $^{15}\text{N}$ – $^1\text{H}$  dipolar coupling and  $^{15}\text{N}$  chemical shift anisotropy (CSA) mechanisms, resulting in very narrow  $^{15}\text{N}$  line widths, a property utilized in the original TROSY-HSQC method for obtaining spectral resolution enhancement in larger proteins (Pervushin et al. 1997). In contrast, the upfield doublet component has increased  $^{15}\text{N}$  line widths relative to a  $^1\text{H}$ -decoupled  $^{15}\text{N}$  resonance, and can become vanishingly weak in larger proteins, making it difficult or impossible to measure the  $^1J_{\text{NH}}$  or  $^1J_{\text{NH}} + ^1D_{\text{NH}}$  splittings. The adverse impact of relaxation interference broadening of the upfield  $^{15}\text{N}$ – $\{^1\text{H}\}$  doublet component on the accuracy of the measured splitting can be mitigated by instead measuring

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N. C. Fitzkee · A. Bax (✉)  
Laboratory of Chemical Physics, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, MD 20892-0520, USA  
e-mail: bax@nih.gov

halved couplings as the splitting between the TROSY component and the peak position in a conventional decoupled  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectrum (Kontaxis et al. 2000). More generally, by limiting the time during which the broad upfield component evolves in the transverse plane, a scaled  $^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}}$  splitting can be measured at a relative accuracy that significantly exceeds that of the unscaled splitting (Yang et al. 1999; Bhattacharya et al. 2010; Mantylahti et al. 2010). An elegant implementation of this principle distributes the relaxation losses equally over the two doublet components, and appears to offer the highest accuracy at which  $^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}}$  splittings can be obtained from frequency domain measurements in larger proteins (Arbogast et al. 2010).

The size of the  $^1\text{J}_{\text{NH}}$  (or  $^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}}$ ) splitting may also be extracted from the relative intensities observed in two or more spectra recorded with pulse schemes where  $^1\text{J}_{\text{NH}}$  dephasing or rephasing is active for different durations,  $\tau$ . Typically, the amplitude of a correlation in the corresponding spectra is a function of  $\sin(\pi\text{J}_{\text{NH}}\tau)$ , and the coupling is extracted from these intensities using the principle of “quantitative J correlation” (Bax et al. 1994). Although intrinsically very precise values can be obtained from such measurements (Tjandra et al. 1996; Tolman and Prestegard 1996), the rapid transverse relaxation of the upfield  $^{15}\text{N}$ - $\{^1\text{H}\}$  doublet component during the  $\tau$  period, caused by relaxation interference between  $^{15}\text{N}$ - $^1\text{H}$  dipolar and  $^{15}\text{N}$  CSA relaxation mechanisms, adversely impacts sensitivity of this approach, in particular for larger proteins.

Here, we introduce an alternate quantitative J-correlation method for  $^1\text{J}_{\text{NH}}$  measurement, where  $^1\text{J}_{\text{NH}}$  dephasing is active while  $^1\text{H}$  magnetization is in the transverse plane, and it is the smaller relaxation interference between  $^1\text{H}^{\text{N}}$  CSA and  $^{15}\text{N}$ - $^1\text{H}$  dipolar coupling which afflicts the measurement. Our new method for measuring amide RDCs by TROSY Spectroscopy is dubbed ARTSY. By limiting the time during which  $^1\text{H}^{\text{N}}$  CSA/ $^{15}\text{N}$ - $^1\text{H}$  dipolar coupling relaxation interference is active to  $(^1\text{J}_{\text{NH}})^{-1} \approx 10.7$  ms, relaxation losses are kept at a minimum, in particular when considering that half of this period already is utilized as part of the standard INEPT transfer (Morris and Freeman 1979) in the regular TROSY-HSQC experiment (Pervushin et al. 1997). By carrying out the measurement on a perdeuterated sample with back-exchanged amide protons, such as often used for NMR studies of larger proteins, such experiments based on  $^1\text{H}$ -dephasing are shown to be remarkably effective for measurement of  $^1\text{D}_{\text{NH}}$  RDCs. We demonstrate the accuracy of ARTSY for the small model protein GB3, and its applicability to larger systems for the catalytic core domain of HIV integrase, a 36 kDa homodimer with rather unfavorable spectral characteristics (Fitzkee et al. 2010).

The ARTSY pulse scheme is very similar to the standard TROSY-HSQC experiment (Fig. 1). The primary differences are that the initial  $^1\text{H}$  dephasing duration of the first INEPT transfer module (between time points *a* and *b*) is set to  $T = (^1\text{J}_{\text{NH}})^{-1} \approx 10.7$  ms, instead of the usual  $(2^1\text{J}_{\text{NH}})^{-1} \approx 5.35$  ms value. The experiment is carried out twice, in an interleaved manner: once with  $^1\text{H}$   $^1\text{J}_{\text{NH}}$  dephasing active during half the *T* interval (reference experiment), and once with  $^1\text{J}_{\text{NH}}$  dephasing active during the full interval (attenuated experiment). The value of the coupling is then extracted from the intensity ratio, *Q*, of the attenuated over the reference intensity:

$$Q = I_A/I_R = \sin[\pi(^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}})T] / \sin[\pi(^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}})T/2] \quad (1)$$

Substituting  $\lambda = ^1\text{J}_{\text{NH}} + ^1\text{D}_{\text{NH}}$  (where  $^1\text{J}_{\text{NH}} \approx -92$  Hz), Eq. 1 can be rewritten as:

$$Q = \sin(\pi\lambda T) / \sin(\pi\lambda T/2) = 2 \cos(\pi\lambda T/2) \quad (2a)$$

or

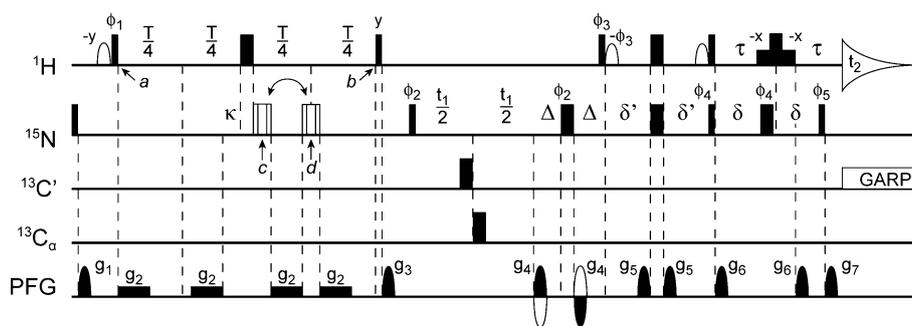
$$\lambda = \pm(2/\pi T) \cos^{-1}(Q/2) \quad (2b)$$

Alternatively, using the trigonometric identity  $\cos^{-1}(x) = \pi/2 - \sin^{-1}(x)$  and the fact that the sign of  $^1\text{J}_{\text{NH}}$  is negative this result can be rewritten as:

$$^1\text{D}_{\text{NH}} = -1/T - ^1\text{J}_{\text{NH}} + (2/\pi T) \sin^{-1}(Q/2) \quad (3)$$

The above analysis ignores relaxation interference between the  $^1\text{H}^{\text{N}}$  CSA tensor and the  $^1\text{H}$ - $^{15}\text{N}$  dipolar interaction, which causes the two  $^1\text{H}$ - $\{^{15}\text{N}\}$  components to decay at different rates,  $R_2 \pm \Gamma$ , during the *T* interval, where  $\Gamma$  is the  $^1\text{H}^{\text{N}}$  CSA/ $^{15}\text{N}$ - $^1\text{H}$  dipolar cross-correlated relaxation rate. Whereas for the attenuated experiment both components decay by the same factor,  $R_2 T$ , for the reference experiment the two components decay by  $(R_2 \pm \Gamma/2)T$ . For  $(\Gamma T/2)^2 \ll 1$ , to a good approximation this cross correlated relaxation does not significantly impact the buildup of antiphase magnetization at time point *b*. For example, even when scaling the average  $\Gamma = 1.58 \text{ s}^{-1}$  value measured for GB3 at  $\tau_c = 3.3$  ns (Yao et al. 2010b) to a protein tumbling at  $\tau_c = 50$  ns, yielding  $\Gamma \approx 24 \text{ s}^{-1}$ , this results in a reference spectrum intensity increase of only 0.8% and therefore has a negligible effect on the extracted RDC.

The precision at which a coupling can be determined depends on the experimental uncertainty in the ratio, *Q*. If equal numbers of scans are recorded for the reference and attenuated spectrum, the error in *Q* is dominated by the uncertainty in  $\sin(\pi\lambda T)$ . The uncertainty in *Q* is found by assuming identical random noise amplitude, *N*, in both the reference and attenuated spectra. Since *Q* is the ratio of two independent intensity measurements, the error propagation formula for division applies:



**Fig. 1** The ARTSY pulse scheme used to measure  $^1\text{H}$ - $^{15}\text{N}$  RDCs. Narrow and wide pulses correspond to  $90^\circ$  and  $180^\circ$  flip angles, respectively. Unless marked, RF phases of all pulses are  $x$ . The  $^1\text{H}$  carrier frequency is set to the water frequency, and  $^{15}\text{N}$ ,  $^{13}\text{C}'$ , and  $^{13}\text{C}^\alpha$  carriers are set to 117, 176, and 56 ppm, respectively. Open  $^1\text{H}$  pulses are  $90^\circ$ , with a shape corresponding to the center lobe of a sinc( $x$ ) function. The last non-selective  $180^\circ$   $^1\text{H}$  pulse is flanked by two low power 1.2-ms rectangular pulses ( $90^\circ$  each) which aid in  $\text{H}_2\text{O}$  signal suppression (Piotto et al. 1992). The  $180^\circ$  proton pulse during this element is offset slightly (by  $\sim 150$   $\mu\text{s}$ ) relative to the  $180^\circ$   $^{15}\text{N}$  pulse to allow inclusion of the  $g_7$  decoding gradient during the second  $\tau$  delay without requiring first order phase correction in the  $^1\text{H}$  dimension. The open composite  $^{15}\text{N}$  pulse is of the  $90_x-210_y-90_x$  type, compensating for both offset and RF inhomogeneity (Freeman et al. 1980), and is applied at time point  $d$  (reference) or  $c$  (attenuated experiment). The delay  $T$  is set to  $\sim 1/J_{\text{NH}}$  or 10.75 ms. For deriving the coupling values, using Eqs. 2 or 3, dephasing during the pulses needs to be taken into account, and a value  $T + \varepsilon$  is used in Eqs. 2 and 3, with  $\varepsilon = (4\pi)H_{90} - \kappa$ , where  $H_{90}$  equals the  $^1\text{H}$   $90^\circ$  pulse width and  $\kappa$  equals the total duration of the  $^{15}\text{N}$  composite pulse.

Other delays:  $\delta = \delta' = \sim 1/4J_{\text{NH}} = 2.71$  ms;  $\Delta = 0.75$  ms;  $\tau = 1.66$  ms. The  $^{13}\text{C}'$  and  $^{13}\text{C}^\alpha$  decoupling pulses (only needed for proteins also enriched in  $^{13}\text{C}$ ) are applied with a RF field strength of  $\Delta f/\sqrt{3}$ , where  $\Delta f$  is the difference in Hz between the centers of the  $^{13}\text{C}'$  and  $^{13}\text{C}^\alpha$  chemical shift regions. Gradients:  $g_2 = 0.7$  G/cm;  $g_{1,3,4,5,6,7}$  are sine-bell shaped with durations of 1.0, 0.5, 0.5, 1.0, 0.8, and 0.1013 ms, respectively, and strengths of 20.3, 8.4,  $\pm 42$ , 2.3, 4.9, and 42 G/cm at their mid-points. Phase cycling:  $\phi_1 = x, -x$ ;  $\phi_2 = 2(y), 2(-y)$ ;  $\phi_3 = -y$ ;  $\phi_4 = 4(x), 4(-x)$ ;  $\phi_5 = 4(-y), 4(y)$ ;  $\phi_{\text{rec}} = y, 2(-y), y, -y, 2(y), -y$ . Phase  $\phi_5$  and the duration of  $\delta'$  may be adjusted to reduce anti-TROSY artifacts according to the Clean TROSY method. Given the average relative intensity of the anti-TROSY artifacts,  $\phi_5$  is generally decreased and  $\delta'$  is shortened according to Eq. 10 of (Schulte-Herbruggen and Sorensen 2000). In practice, anti-TROSY artifacts were vanishingly weak in our integrase spectra, and therefore no such compensation was used. Quadrature in the  $^{15}\text{N}$  dimension uses the regular gradient coherence selection method, inverting the phases of  $\phi_3, \phi_5$  and the sign of  $g_4$  for the second FID collected for each value of  $t_1$  (Kay et al. 1992; Pervushin et al. 1998)

$$\sigma_Q = \frac{|I_A|}{|I_R|} \sqrt{\left(\frac{N}{I_R}\right)^2 + \left(\frac{N}{I_A}\right)^2} = \sqrt{Q^2 + 1}/(I_R/N) \quad (4a)$$

where  $I_R$  and  $I_A$  denote the respective intensities in the reference and attenuated spectra. Once again, using standard error propagation, the uncertainty in  $\lambda$  is then given by:

$$\sigma_\lambda = \sigma_Q \frac{\partial}{\partial Q} \lambda(Q) \Big|_Q = \frac{2}{\pi T (I_R/N)} \sqrt{\frac{1+Q^2}{4-Q^2}} \quad (4b)$$

For the common situation where  $Q \ll 1$ , Eq. 4 simplifies to  $\sigma_\lambda \approx 1/[\pi T(S/N)]$ . This means that for the recommended  $T = 10.7$  ms, and an observed  $Q \sim 0$ , a  $S/N$  ratio of 30 in the reference spectrum yields a noise-limited uncertainty in the derived coupling of ca. 1 Hz, more than sufficient for most applications to larger proteins.

In practice, the value for  $T$  is adjusted such as to maximize sensitivity of the measured couplings, which occurs for  $Q \approx 0$ , i.e.,  $T = N/|J_{\text{NH}} + D_{\text{NH}}|$ . If the  $^1J_{\text{NH}} + D_{\text{NH}}$  values span a narrow range, a value for  $N$  chosen such that  $T \approx T_2$ , where  $T_2$  is the  $^1\text{H}^{\text{N}}$  transverse relaxation time, will yield the highest precision for the extracted coupling. For application to perdeuterated proteins where alignment strength is typically relatively large ( $D_{\text{NH}}$  in the  $\pm 25$  Hz

range) and the  $^1\text{H}^{\text{N}}$   $T_2$  is short, the optimum  $T$  value is obtained for  $N = 1$ . With  $D_{\text{NH}}$  values roughly equally distributed about zero, this then requires  $T \approx 1/|J_{\text{NH}}| \approx 10.7$  ms.

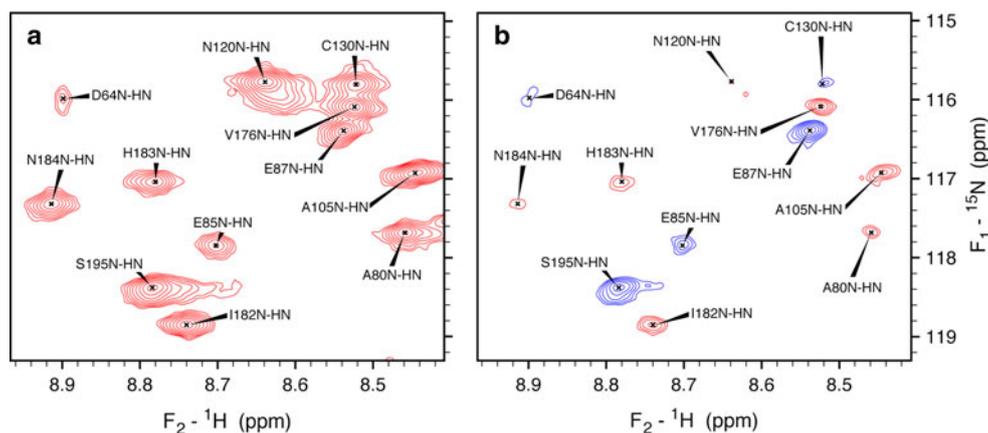
Accuracy of the method is demonstrated for the perdeuterated, amide-protonated model system GB3, either in isotropic buffer or dissolved in a liquid crystalline Pf1 suspension (11 mg/ml). With a pairwise root-mean-square difference (rmsd) of 0.19 Hz, isotropic  $^1J_{\text{NH}}$  values agree well with values previously measured with a gradient-enhanced IPAP-HSQC experiment (Yao et al. 2009) (Supporting Information). Similarly, with a Pearson's correlation coefficient of  $R_P = 0.996$ , RDCs derived from the difference in  $^1J_{\text{NH}} + D_{\text{NH}}$  values in aligned and isotropic samples fit very well to the RDC-refined structure of GB3 (PDB entry 2OED (Ulmer et al. 2003); Supporting Information).

As an application to a system of biological interest, we used ARTSY to measure  $D_{\text{NH}}$  RDC values in a sample of the catalytic core domain of the HIV-1 integrase enzyme. The domain studied comprises residues 50–212, and below is referred to as  $\text{IN}^{50-212}$ . It includes the following mutations, introduced to improve its solubility and stability: Q53E C56S W131E F185 K Q209E. Even with these mutations, this homodimeric protein (36 kDa) has rather

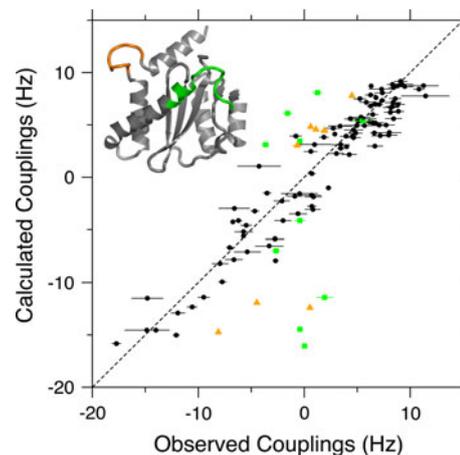
unfavorable NMR spectral characteristics and is of limited stability. Its NMR assignment and preliminary characterization was reported recently (Fitzkee et al. 2010), and both chemical shift analysis and  $^1\text{H}^{\text{N}}\text{-}^1\text{H}^{\text{N}}$  NOEs measured on the perdeuterated protein indicated that the protein in solution adopts a structure very similar to that observed in the crystalline state (Dyda et al. 1994; Bujacz et al. 1996; Goldgur et al. 1998; Chen et al. 2000). The N-terminal tail, the active site loop, and the loop preceding the final  $\alpha$ -helix, for which electron density was missing in many of the X-ray structures, were found to be dynamically disordered as judged by  $^{15}\text{N}$  relaxation data (Fitzkee et al. 2010).

ARTSY data for a sample containing 0.4 mM of  $\text{IN}^{50-212}$  (0.2 mM dimer) in 4% (w/v) liquid crystalline C12E5 poly-ethylene-glycol/hexanol mixture ( $^2\text{H}$  splitting = 19.6 Hz) (Ruckert and Otting 2000) were recorded on a Bruker Avance 900 MHz spectrometer, equipped with a cryogenic probe. The data were recorded at 298 K in a 20 mM PIPES pH 6.6 buffer, containing 150 mM NaCl and 40 mM  $\text{MgCl}_2$ . The sample was prepared as described previously (Fitzkee et al. 2010). Interleaved reference and attenuated spectra were recorded with acquisition times of 80 ms ( $t_1$ , 250 complex points), and 120 ms ( $t_2$ , 1,784 complex points) and a recycle delay of 1.5 s, with 64 scans per recorded transient, and a total acquisition time of 27 h.

Figure 2 compares a small region of the  $\text{IN}^{50-212}$  ARTSY reference spectrum with that of the attenuated spectrum, and the vast majority of previously assigned  $^1\text{H}\text{-}^{15}\text{N}$  correlations (Fitzkee et al. 2010) is observed in the reference spectrum. In the attenuated spectrum (Fig. 2b), amides with  $|^1J_{\text{NH}} + ^1D_{\text{NH}}| > 93$  Hz are opposite in sign relative to those with smaller values. A nearly complete set of RDCs could be extracted from these spectra (Supporting



**Fig. 2** Spectral data used for measuring  $^1\text{H}\text{-}^{15}\text{N}$  RDCs in  $\text{IN}^{50-212}$ . Small regions of the reference (a) and attenuated (b)  $^{15}\text{N}\text{-}^1\text{H}$  TROSY spectra, recorded at 900 MHz in the presence of 4% (w/v) C12E5 PEG/hexanol (Ruckert and Otting 2000). Peak positions are determined using the reference spectra and then mapped directly onto the



**Fig. 3** Best fit correlation between ARTSY-derived  $^1D_{\text{NH}}$  RDCs for homodimeric  $\text{IN}^{50-192}$  and the coordinates of PDB entry 1BIS, chain B (Goldgur et al. 1998). The alignment tensor was best-fitted using the DC program in the NMRPipe software package (Delaglio et al. 1995). Black symbols correspond to the 97 RDCs used in fitting ( $Q = 0.279$ ). Green squares and orange triangles correspond to the active site loop (residues 140–152) and the loop between helices  $\alpha_5$  and  $\alpha_6$  (residues 187–194) (see colored regions on the inset structure) and were not included in the RDC fit. These amides were shown to be dynamic in  $^{15}\text{N}$  relaxation measurements (Fitzkee et al. 2010). Slightly better fits are obtained using the chains of PDB entry 1BIU ( $Q \sim 0.26$ ), but its chains lack density for residues 141–148 of the catalytic loop (Goldgur et al. 1998)

Information). Fitting of these RDCs to the various crystal structures shows fair agreement. For example, chain B of PDB entry 1BIS (Goldgur et al. 1998) exhibits a  $Q$  factor of 0.28 (Fig. 3), only slightly above the value of 0.2–0.25 expected for a protein solved at 2-Å resolution (Bax 2003). This chain includes coordinates for all of the catalytic loop residues (140–152) as well as for the loop (residues 187–194) immediately preceding the  $\alpha_6$  helix. However, as

attenuated spectra. Positive and negative intensity contours are red and blue, respectively. In the attenuated spectra, negative (positive) peaks correspond to resonances with  $|J + D|$  greater than (less than) 93 Hz

seen in Fig. 3, the active site loop fits very poorly to the measured RDCs. The near-zero RDC values observed for most of the active site amide groups are consistent with a high degree of internal dynamics, as also reflected in missing electron density in many other integrase crystal structures (Dyda et al. 1994; Bujacz et al. 1996; Chen et al. 2000) and our previous  $^{15}\text{N}$  relaxation measurements.

ARTSY is particularly well suited for proteins that have poor spectral characteristics, such as the HIV1 integrase catalytic core domain dimer, but the same idea is of course also applicable to less challenging systems too. For smaller proteins, where use of TROSY is not essential, the same ARTSY  $^1\text{J}_{\text{NH}}$  amplitude encoding can be used in the regular gradient-enhanced  $^1\text{H}$ – $^{15}\text{N}$  HSQC experiment (Kay et al. 1992). Similarly, for proteins with substantial resonance overlap in the 2D HSQC or TROSY-HSQC spectra, the ARTSY  $^1\text{J}_{\text{NH}}$  amplitude encoding can be used during the initial  $^1\text{H} \rightarrow ^{15}\text{N}$  INEPT transfer of better resolved 3D experiments, such as HNC0, or HNC0-TROSY, albeit with the concomitant loss associated with carrying out the experiments in a 3D rather than a 2D fashion. Even for the 723-residue protein malate synthase G, an HNC0-TROSY allowed sufficient resolution for measuring more than 95% of the  $^1\text{H}$ – $^{15}\text{N}$  RDCs using the J-scaling approach (Tugarinov and Kay 2003). Thus, ARTSY should be particularly useful for even more challenging systems, as no resolution is sacrificed to measure the couplings.

One additional point requiring attention is the application of ARTSY to protonated proteins. Although in principle, ARTSY is applicable to protonated proteins, for the TROSY version of the experiment a complicating factor is caused by the buildup of antiphase terms of the type  $\text{H}_x^{\text{J}}\text{N}_z$  between time points  $a$  and  $b$  in Fig. 1, where  $\text{H}^{\text{J}}$  denotes protons with multi-bond  $\text{J}_{\text{NH}}$  couplings to the  $^{15}\text{N}$  considered. These terms transfer into  $\text{N}_x\text{H}_z^{\text{J}}$  at the start of  $^{15}\text{N}$  evolution (Supporting Information). Although in regular HSQC experiments, such terms yield vanishing intensity as they remain antiphase with respect to  $\text{H}^{\text{J}}$  during  $t_1$  evolution and therefore do not result in detectable in-phase  $^1\text{H}$  magnetization during detection, in TROSY-HSQC experiments these terms partially rephase during  $t_1$  evolution, giving rise to unresolved anti-phase doublets in the  $^{15}\text{N}$  dimension of the spectrum (Supporting Information). Provided that the peak position at which the intensity in the attenuated spectrum is measured is taken from that determined in the reference spectrum (a feature of the NMRPipe (Delaglio et al. 1995) seriesTab peak picking procedure used in our study), the antiphase component does not contribute to the intensity measured in the attenuated spectrum,  $I_A$ . Additionally, aliphatic protons can be decoupled by using an amide proton selective pulse during the initial INEPT period, and this will largely eliminate the buildup of antiphase terms (Supporting Information).

Indeed, ARTSY-derived RDCs for fully protonated GB3 fit very well to the experimentally determined (Yao et al. 2010a) GB3 bond vectors ( $R_p = 0.998$ ; Supporting Information). Although, at least in principle, unresolved long range  $^1\text{H}^{\text{N}}$ – $^{15}\text{N}$  RDCs could result in similar antiphase contributions to the  $^{15}\text{N}$  line shape in ARTSY spectra on perdeuterated proteins, in practice long range  $^1\text{H}^{\text{N}}$ – $^{15}\text{N}$  RDCs tend to be very small, and no such antiphase contributions were detected for the  $\text{IN}^{50-212}$  sample. As found for protonated GB3, even if such small antiphase contributions were present, to first order they do not impact the  $I_A/I_R$  intensity ratios, used for deriving the coupling values.

Base line correction is one important detail requiring special attention when applying ARTSY to larger proteins that also contain highly disordered residues, giving rise to very intense resonances. It therefore is recommended to record the spectra in such a manner that no frequency dependent linear baseline correction is needed, while scaling the first data point of each FID (particularly in the  $t_1$  dimension) by 0.5 (Otting et al. 1986), or by recording the spectra such that a  $180^\circ$  linear phase correction is needed with no scaling of the first data point (Zhu et al. 1993).

In summary, ARTSY is an effective alternative for recording amide  $^1\text{H}$ – $^{15}\text{N}$  RDCs, particularly well suited for larger perdeuterated proteins. The spectral resolution is identical to that of the regular TROSY-HSQC spectrum, which greatly benefits from using the highest available magnetic field strength. Noise in the reference spectrum impacts the size of the coupling, extracted from Eq. 2, far less than noise in the attenuated spectrum. In principle, therefore, it is favorable to record the attenuated spectrum with more scans than the reference spectrum, and correspondingly scale the ratio  $Q$  before applying Eqs. 2b or 3. In practice, sensitivity of the spectra recorded for  $\text{IN}^{50-212}$  was more than sufficient for accurate measurement of  $^1\text{D}_{\text{NH}}$  RDCs, and the use of equal numbers of scans for both reference and attenuated spectra was adequate.

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