

Thermodynamic Rearrangement Synthesis and NMR Structures of C_1 , C_3 , and T Isomers of $C_{60}H_{36}$

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Abstract: The structures of three $C_{60}H_{36}$ isomers, produced by high-temperature transfer hydrogenation of C_{60} in a 9,10-dihydroanthracene melt, was accomplished by 2D 1H -detected NMR experiments, recorded at 800 MHz. The unsymmetrical C_1 isomer is found to be the most abundant one (60–70%), followed by the C_3 isomer (25–30%) and the least abundant T isomer (2–5%). All three isomers are closely related in structure and have three vicinal hydrogens located on each of the 12 pentagons. Facile hydrogen migration on the fullerene surface during annealing at elevated temperatures is believed to be responsible for the preferential formation of these thermodynamically most stable $C_{60}H_{36}$ isomers. This hypothesis was further supported by thermal conversion of $C_{60}H_{36}$ isomers to a single C_{3v} isomer of $C_{60}H_{18}$.

Introduction

$C_{60}H_{36}$ was one of the first derivatives of C_{60} ever made,¹ and it appears to be the ultimate hydrogenation product under various hydrogenation conditions.^{2,3} This and other hydrofullerenes are of particular interest as models for the investigation of surface hydrogenation–dehydrogenation processes in carbon nanosystems. Fully surface-hydrogenated carbon nanomaterials can reach hydrogen capacity as high as 7.7% for $CH_{1.0}$ composition, thus exceeding the 6.5% hydrogen storage systems requirement.^{3,4}

Despite its easy availability and prodigious interest, $C_{60}H_{36}$ has defied numerous attempts to determine its structure for almost 10 years. Selected bibliographic materials with 67 references pertaining to the history of $C_{60}H_{36}$ research are included as Supporting Information. Here, we report the results of our NMR structure determination of the three predominant $C_{60}H_{36}$ isomers, consistently produced by high-temperature hydrogenation of C_{60} , which provide compelling evidence for facile hydrogen migration on the fullerene surface at high temperatures.

Results and Discussion

Hydrogenation of C_{60} to $C_{60}H_{36}$ can be performed by several methods, including Birch hydrogenation, catalytic hydrogenation, Zn-alloy reduction in HCl, and radical transfer hydrogenation.^{3,4}

A recent report indicated that low-temperature hydrogenation (Birch conditions) affords a complex mixture of isomers, whereas high-temperature transfer hydrogenation in a 9,10-dihydroanthracene melt at 350 °C yields mainly two isomers of $C_{60}H_{36}$ (C_1 and C_3), in roughly a 3:1 ratio.⁵ Because of this apparent isomer selectivity, we chose this method to generate material for structural exploration.

Synthesis. High-temperature hydrogenation of $C_{60}H_{36}$ was performed as described earlier,^{5,6} but with some minor modifications. The reported procedure⁵ involved HPLC purification of the products. To avoid potential loss and/or isomerization of $C_{60}H_{36}$ isomers during HPLC separation, we investigated this reaction in an attempt to find conditions at which $C_{60}H_{36}$ samples can be obtained with satisfactory purity for NMR analysis without requiring chromatographic separation. The major factors influencing the production of high-quality $C_{60}H_{36}$ samples were found to be time and temperature. Longer reaction time (1.5 h compared to 0.75 h at 350 °C)⁵ favored isomer selectivity (annealing process, see below). Unfortunately, it also promoted subsequent dehydrogenation of $C_{60}H_{36}$ isomers to C_{3v} $C_{60}H_{18}$.^{6,7} The best results were achieved when the reaction was carried out at slightly lower temperature, 340 ± 10 °C, for a total duration of 2 h. The $C_{60}H_{36}$ samples had satisfactory stability in the solution (4–6 h), even in the presence of a small amount of oxygen. They also had reasonable solubility in $CDCl_3$ (5–7 g/L), thus eliminating the need of toxic CS_2 as a cosolvent.

Structure Determination. The very limited spectral dispersion in both the 1H and ^{13}C spectra of the mixture of $C_{60}H_{36}$

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- (1) Hauffler, R. E.; Conceicao, J.; Chibante, L. P. F.; Chai, Y.; Byrne, N. E.; Flanagan, S.; Haley, M. M.; O'Brien, S. C.; Pan, C.; Xiao, Z.; Billups, W. E.; Ciufolini, M. A.; Hauge, R. H.; Margrave, J. L.; Wilson, L. J.; Curl, R. F.; Smalley, R. E. *J. Phys. Chem.* **1990**, *94*, 8634–8636.
- (2) Nossal, J.; Saini, R. K.; Alemany, L. B.; Meier, M.; Billups, W. E. *Eur. J. Org. Chem.* **2001**, 4167–4180.
- (3) Tarasov, B. P.; Goldshleger, N. F.; Moravskii, A. P. *Usp. Khim.* **2001**, *70*, 149–166.
- (4) Dillion, A. C.; Heben, M. J. *Appl. Phys. A* **2001**, *72*, 133–142.

- (5) Nossal, J.; Saini, R. K.; Sadana, A. K.; Bettinger, H. F.; Alemany, L. B.; Scuseria, G. E.; Billups, W. E.; Saunders, M.; Khong, A.; Weisemann, R. *J. Am. Chem. Soc.* **2001**, *123*, 8482–8495.
- (6) Rüdhardt, C.; Gerst, M.; Ebenhoch, J.; Beckhaus, H.-D.; Campbell, E. E. B.; Tellmann, R.; Schwarz, H.; Weiske, T.; Pitter, S. *Angew. Chem., Int. Ed. Engl.* **1993**, *32*, 584–586.
- (7) Darvish, A. D.; Avent, A. G.; Taylor, R.; Walton, D. R. M. *J. Chem. Soc., Perkin Trans. 2* **1996**, 2051–2054.

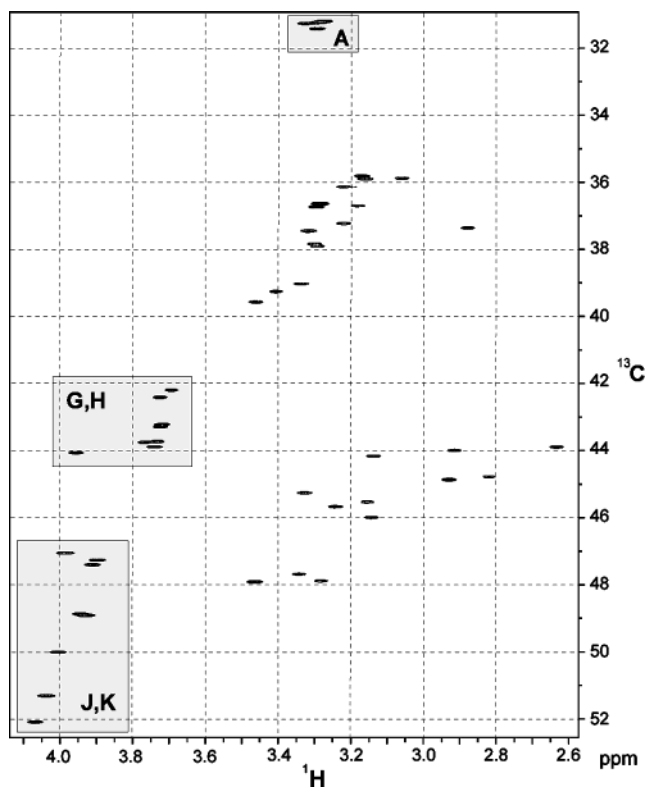


Figure 1. The 800-MHz HSQC spectrum of C_1 , C_3 , and T $C_{60}H_{36}$ mixture.

isomers required both the highest available magnetic field strength and special adaptation of the inverse detected NMR experiments, to obtain sufficient resolution in the inverse detected 1H - ^{13}C correlation spectra. Because of the long ^{13}C transverse relaxation times of $C_{60}H_{36}$ resonances in $CDCl_3$ (> 100 ms), a very large number of increments could be sampled in this dimension without loss of sensitivity per unit of measuring time.⁸ This resulted in excellent resolution in the ^{13}C dimension, with line widths of about 0.03 ppm. Only three 2D NMR spectra were utilized for the structure determination process: 1H - ^{13}C HSQC⁹ for identifying the one-bond correlations (Figure 1), 1H - ^{13}C COSY-HSQC¹⁰ for connecting ^{13}C resonances to protons two-bonds removed, and 1H - ^{13}C TOCSY-HSQC¹⁰ for resolving ambiguities in the analysis of the 1H - ^{13}C COSY-HSQC spectrum. The high resolution obtained in the ^{13}C dimension was critical for minimizing overlap, particularly in the relayed 2D spectra.

^{13}C and 1H spectral data of $C_{60}H_{36}$ samples (CH fragments) are presented in Table 1 and are in general agreement with the previously reported data.⁵ The spectra have 36 signals of CH fragments assigned to a C_1 isomer (60–70%), 12 signals assigned to a C_3 isomer (25–30%), and three additional signals with lower intensity which we assigned to a T isomer (2–5%) of $C_{60}H_{36}$. The assignment of these ^{13}C peak sets was made based on the results of the COSY-HSQC and TOCSY-HSQC experiments and was further confirmed by evaluation of peak heights (cf. A4, A3, A2 with A1; H25 with H24; G22 with G23; K43 with K44, Figure 2).¹¹

Structure elucidation of the unsymmetrical C_1 isomer of $C_{60}H_{36}$ is a formidable task, taking into account the possibility

Table 1. ^{13}C and 1H Chemical Shifts of the Three $C_{60}H_{36}$ Isomers in $CDCl_3$

signal	δ (^{13}C) ^a	δ (1H) ^a	signal	δ (^{13}C) ^a	δ (1H) ^a
C_1 Isomer					
A2	31.34	3.28	G22	43.32	3.72
A3	31.37	3.32	G29	44.17	3.96
A4	31.52	3.29	H21	42.52	3.73
B40	47.79	3.34	H25	43.85	3.77
B41	47.99	3.29	H26	43.99	3.74
C14	37.94	3.30	I30	44.27	3.14
C15	38.01	3.29	31	44.89	2.82
C18	39.36	3.41	I34	45.64	3.16
D16	37.47	2.88	I36	46.10	3.15
D27	44.00	2.63	J37	47.16	3.99
D28	44.10	2.92	J45	50.11	4.01
E6	35.98	3.06	J47	52.20	4.07
E10	36.81	3.18	K38	47.37	3.89
E19	39.67	3.46	K39	47.51	3.91
F7	36.00	3.16	K43	49.01	3.93
F9a	36.74	3.29	L8	36.25	3.22
F11	36.84	3.30	L17	39.13	3.34
G20	42.30	3.69	L33	45.36	3.33
C_3 Isomer					
A1	31.31	3.27	G23	43.39	3.73
B42	48.02	3.47	H24	43.83	3.73
C13	37.56	3.32	I35	45.78	3.24
D32	44.97	2.93	J46	51.42	4.04
E12	37.33	3.22	K44	49.02	3.92
F5	35.92	3.17	L9b	36.75	3.29
T Isomer					
A	31.37	3.19			
H	43.61	3.67			
K	49.25	3.86			

^a NMR data are presented in ppm versus internal reference [$CDCl_3$ at δ 77.23 ppm (^{13}C) and 7.24 (1H)].

of more than 600 trillion solutions.¹² The key to solving the structure was the identification of six CH groups interconnected in a six-member ring. Only one such combination was found: A2-K39-A3-K38-A4-K43 (Figure 3). There exists only one possible placement for this ring in the C_{60} framework (Figure 4a). The discovery of this key fragment allows for step-by-step structure solution (see detailed discussion in the Supporting Information).

Unique assignment of all 36 CH groups in the structure of the C_1 isomer of $C_{60}H_{36}$ provided an opportunity for 2D area analysis (^{13}C , 1H) of their NMR signals. Several compact groups of signals were identified (Figure 1). Type A signals (A2, A3, A4) were in a very narrow range of 31.3–31.6/3.27–3.32 ppm ^{13}C / 1H . Other compact groups of signals were J/K types in the 47.1–52.2/3.89–4.07 ppm region and G/H types, all located within the narrow range of 42.2–44.2/3.69–3.96 ppm.

Structure analysis of the C_3 isomer is initiated by identification of the signals of three identical $CHCHCHCHCH(CH)_2$ fragments as A1-K44-H24-G23-J46(C13,F5) in the COSY-HSQC spectrum. Due to the C_3 symmetry, there is only one possible placement of these three fragments in the C_{60} framework (Figure 4b). A detailed description of C_3 structure elucidation is included as Supporting Information.

(11) In our $C_{60}H_{36}$ samples the C_3/C_1 ratio was slightly higher than the exact 1/3 ratio reported previously.⁵ As a result, C_3 signals (magnified three times compared to C_1 signals due to the symmetry) were about 10–20% higher than the corresponding C_1 signals. The T isomer signal set was identified on the basis of its high symmetry, but lower relative abundance (2–5%), which translates to about half the relative intensity compared to signals of the C_1 isomer (see Figure 2). Smaller signal intensities of the T isomer have been observed in the $CDCl_3$ saturated solutions used for recording the 2D NMR spectra. Reference spectra of C_1/C_3 mixtures without the T isomer were reported elsewhere.⁵

(12) Balasubramanian, K. *Chem. Phys. Lett.* **1991**, *182*, 257–262.

(8) Levitt, M. H.; Bodenhausen, G.; Ernst, R. R. *J. Magn. Reson.* **1984**, *58* (3), 462–472.

(9) Bodenhausen, G.; Ruben, D. J. *Chem. Phys. Lett.* **1980**, *69*, 185–189.

(10) Lerner, L.; Bax, A. *Carbohydr. Res.* **1987**, *166*, 35–46.

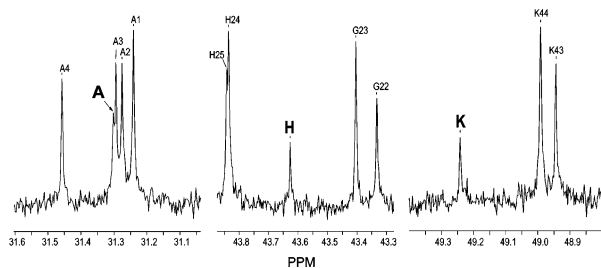


Figure 2. ^{13}C spectra showing three peaks (A, H, K) of the *T* isomer of $\text{C}_{60}\text{H}_{36}$.

Structure determination of the elusive⁵ *T* isomer of $\text{C}_{60}\text{H}_{36}$ was made by using the results of 2D NMR experiments and computer analysis (see the Supporting Information). Final selection was made by calculation of relative thermodynamic stabilities of the only two candidates compatible with the NMR data (a second *T* isomer is almost 50 kcal/mol less thermodynamically stable than the selected one, Figure 4c). Similar computational results have been reported previously.¹³

Surface Hydrogenation. Determination of the structures of all three $\text{C}_{60}\text{H}_{36}$ isomers (together comprising over 80% of the $\text{C}_{60}\text{H}_{36}$ samples) provides additional insight into the nature of surface hydrogenation of the fullerene C_{60} and the structure of these hydrides.

First, all three isomers are closely related in structure as well as in their “tetrahedral” 3D appearance (Figure 4d). The most abundant C_1 isomer and less abundant C_3 isomer differ only by the position of one isolated double bond, which has little effect on the overall geometry of the molecule. In the minor *T* isomer, three out of four benzene fragments are positioned identically to both C_1 and C_3 isomers.

It is also notable that in all three isomers, pentagons have exactly three vicinal hydrogen atoms attached. This explains the “magic” substitution number (36): the original C_{60} structure contains 12 pentagons. The underlying reasons for this behavior are not clear at this time, but thermodynamic considerations must play a major role. Computer calculations showed that displacement of one isolated double bond to a hexagon–hexagon junction from a pentagon–hexagon juncture results in a substantial energy loss.¹³ While it is premature to conclude that all $\text{C}_{60}\text{H}_{36}$ isomers would adopt this rule, thus far all known structures of $\text{C}_{60}\text{X}_{36-48}$ ($X = \text{H}, \text{F}$) seem to obey it.^{14–16}

Computational modeling demonstrated another strong link between the three $\text{C}_{60}\text{H}_{36}$ isomers reported here: they all belong to a set of the thermodynamically most stable structures.^{5,13} Unfortunately, the precision of existing computational methods does not allow for a clear distinction between $\text{C}_{60}\text{H}_{36}$ isomers (the differences in thermodynamic stability were typically less than 0.2 kcal/bond and therefore statistically indistinguishable).⁵ Nevertheless, certain trends are clear from the analysis of these data.

The average calculated thermodynamic stabilities of the *T*, C_3 , and C_1 isomers rank as follows: $T \sim C_3 > C_1$.^{5,13} The actual

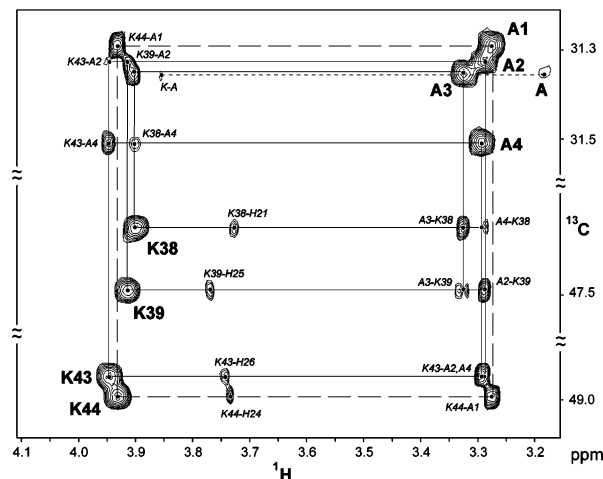


Figure 3. Composite region of the COSY-HSQC spectrum, showing the key connectivities for the six-membered protonated ring of the C_1 isomer (solid lines), the C_3 isomer (long dashes), and the *T* isomer (short dashes). One-bond ^1H – ^{13}C connectivities are marked in regular font. Cross-peaks are marked in italic type. The spectrum has been phased so that the diagonal peaks (one-bond connectivities) are absorptive, and cross-peaks (two-bond connectivities) are antiphase dispersive in the ^1H dimension. Only positive contours are shown.

distribution of these isomers in $\text{C}_{60}\text{H}_{36}$ samples is almost reversed: $C_1 > C_3 \gg T$. Although the existence of residual kinetic factors cannot be ruled out,¹⁷ the experimental data regarding high-temperature hydrogenation of C_{60} argue against this possibility. A plausible explanation of this reverse distribution pattern involves consideration of the entropy component in the Gibb’s free energy equation which governs the equilibrated processes: $\Delta G = \Delta H - T\Delta S$. The entropy factor would favor the formation of the least symmetrical isomers (other factors being equal), which is in agreement with the experimental data.

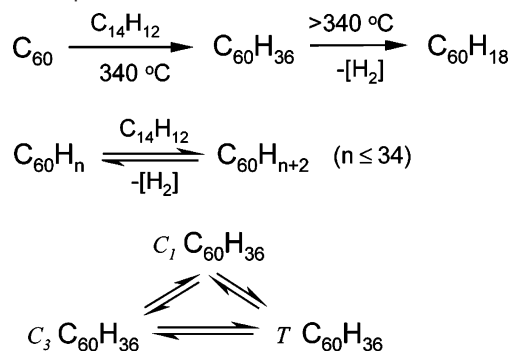
The computer modeling studies also provided some additional insight regarding the observed stoichiometric selectivity of the hydrogenation process. As mentioned earlier, $\text{C}_{60}\text{H}_{36}$ is the ultimate product of many hydrogenation reactions. Computations showed that less hydrogenated molecules, such as $\text{C}_{60}\text{H}_{34}$, typically have less (or about the same) strain energy compared to that of $\text{C}_{60}\text{H}_{36}$. At the same time, more hydrogenated molecules, such as $\text{C}_{60}\text{H}_{38}$ and higher, are significantly more strained. The likely reason for this is unfavorable hydrogen–hydrogen interactions in highly hydrogenated pentagons. Structural constraints dictate that starting with $\text{C}_{60}\text{H}_{38}$ all hydro[60]-fullerenes contain five-membered rings with at least four hydrogen atoms. Similar computational results were reported for $\text{C}_{70}\text{H}_{36}$, suggesting the general nature of these observations.¹⁸

Hydrogen Rearrangement. Available experimental data (primarily improvement in isomer selectivity by applying “annealing” for the preparation of $\text{C}_{60}\text{H}_{36}$, as well as thermodynamic properties of thus produced isomers) point to the

- (13) Clare, B. W.; Kepert, D. L. *J. Mol. Struct.* **1999**, *466*, 177–186. Clare, B. W.; Kepert, D. L. *J. Mol. Struct.* **2002**, *589*, 195–207.
 (14) Gakh, A. A.; Tuinman, A. A.; Adcock, J. L.; Sachleben, R. A.; Compton, R. N. *J. Am. Chem. Soc.* **1994**, *116*, 819–820.
 (15) Gakh, A. A.; Tuinman, A. A. *Tetrahedron Lett.* **2001**, *42* (41), 7133–7135.
 (16) Avent, A. G.; Clare, B. W.; Hitchcock, P. B.; Kepert, D. L.; Taylor, R. *Chem. Commun.* **2002**, (20), 2370–2371.

- (17) An alternative explanation of the “reverse” distribution of isomers entails slow kinetics of the interconversion of C_1 , C_3 , and *T* isomers compared to fast isomerization of other less thermodynamically stable isomers to C_1 , C_3 , and *T* mixture. Although the $T/C_3/C_1$ ratios improve somehow in samples that were prepared at higher temperatures and/or longer reaction times, these 10–30% differences cannot fully explain the observed anomaly. See also: Gakh, A. A.; Tuinman, A. A. *Tetrahedron Lett.* **2001**, *42* (41), 7137–7139.
 (18) Gerst, M.; Beckhaus, H.-D.; Rüchardt, C.; Campbell, E. E. B.; Tellmann, R. *Tetrahedron Lett.* **1993**, *34* (48), 7729–7732.

Scheme 1. Hydrogenation/Dehydrogenation¹⁹/Isomerization Patterns in Fullerene C₆₀–9,10-Dihydroanthracene Mixtures at Elevated Temperatures^a



^a The actual equilibrium may also include other C₆₀H₃₆ isomers (not shown).

of existing experimental results (such as the three vicinal hydrogens per pentagon rule for C₆₀H₃₆ and related systems), an assumption can be made that hydrogenation of all-hexagon carbon nanostructures would follow certain structural patterns. It is also possible that hydrogenation at elevated temperatures favors facile hydrogen migration, leading to tight surface packing of hydrogen atoms and to production of materials with high mass content of usable hydrogen.

Experimental Section

NMR Spectroscopy. Two-dimensional NMR spectra were recorded at 800 MHz, 20 °C, on saturated solutions of C₆₀H₃₆ in CDCl₃ (approximately 5 mg/mL): a regular inverse [¹H,¹³C] HSQC spectrum;²⁰ an inverse COSY-HSQC spectrum with a 20 ms constant-time COSY delay;¹⁰ a TOCSY-HSQC spectrum,¹⁰ recorded with a 18 ms ¹H DIPSI-2 mixing²¹ duration using a 10 kHz RF field strength. The HSQC and TOCSY-HSQC spectra were recorded with gradient enhancement.²² All spectra were recorded with a 10 kHz (49.7 ppm) spectral width in the indirect (¹³C) dimension, and a 5.5 ppm spectral width in the ¹H dimension, using 16 scans and total acquisition times of 12 h (HSQC) and 16 h (COSY-HSQC and TOCSY-HSQC). Acquired data matrices consisted of 800* × 512* data points (HSQC) and 1200* × 512* data points (COSY-HSQC and TOCSY-HSQC). Time domain data were apodized with 80°-shifted sine bell and squared sine bell windows in the *t*₁ and *t*₂ dimensions, in both cases truncated at 170°. Data matrices

(20) Bodenhausen, G.; Rueben, D. J. *Chem. Phys. Lett.* **1980**, *69*, 185–189.

(21) Shaka, A. J.; Lee, C. J.; Pines, A. *J. Magn. Reson.* **1988**, *77*, 274–293.

(22) Kay, L. E.; Keifer, P.; Saarinen, T. *J. Am. Chem. Soc.* **1992**, *114*, 10663–10665.

were zero-filled to yield a digital resolution of 2.5 Hz (¹³C) and 4.3 Hz (¹H). Data were processed and analyzed using the NMRPipe software package.²³ One-dimensional NMR spectra were recorded at 400 MHz, 23 °C, on solutions of C₆₀H₃₆ in CS₂/CDCl₃ (1:1).

Molecular Calculations. The geometries and relative thermodynamic characteristics of C₆₀H₃₆ isomers were analyzed using commercially available Chem3D (CambridgeSoft Corporation), Alchemy (Tripos), and HyperChem (Hypercube) computational packages, employing AM1, MM1, MM2, PM3, and 6-31G methods.

Synthesis of C₁, C₃, and T Isomers of C₆₀H₃₆. C₆₀ (200 mg, 0.28 mmol) and 9,10-dihydroanthracene (5.0 g, 28 mmol) were slowly brought to boiling (Wood alloy bath) in a flow of nitrogen. The purple-red mixture was kept for approximately 2 h at 340 ± 10 °C under a slight nitrogen pressure (to prevent excessive boiling of 9,10-dihydroanthracene, bp 312 °C). Progress of the reaction was monitored by TLC (hexane–benzene, hexanes–ethyl acetate). At the end of the reaction excess pressure of nitrogen was released, and the pale yellow reaction mixture was allowed to cool to room temperature. The melt was crushed with a spatula and extracted many times with ether, first by decantation, and then using a Millipore teflon filter. The ether filtrate was periodically checked for the presence of anthracene-derived products by evaporation of a small portion of the solvent and detection of the fluorescence of the residue under a UV lamp. After the disappearance of detectable fluorescence from the filtrate, pale yellow chunks of C₆₀H₃₆ were collected from the filter and dried in vacuo. The compound was dissolved in a minimal quantity of warm benzene, filtered from small amounts of insoluble products, evaporated, and then dried in vacuo to yield 110 mg (0.15 mmol, 52%) of C₆₀H₃₆ as a pale yellowish cream powder. The NMR solutions were filtered before the experiments. The C₆₀H₃₆ samples contained small amounts of benzene even after 12 h of drying in vacuo at elevated temperatures.

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Supporting Information Available: Description of the structure elucidation of the C₁, C₃, and T isomers; description of compatibility/incompatibility studies of C_{3v} C₆₀H₁₈ versus C₁, C₃, and T isomers of C₆₀H₃₆; bibliographic references pertaining to the history of C₆₀H₃₆ research (67 references). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(23) Delaglio, F.; Grzesiek, S.; Vuister, G. W.; Zhu, G.; Pfeifer, J.; Bax, A. *J. Biomol. NMR* **1995**, *6*, 277–293.